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Letter to the Editor

I would like to thank each and everyone who attended and/or helped in any way at our hydrilla workshop/ field days in June and July at Camp Mack. Both days were a tremendous success, as we have already had many positive phone calls, newspaper articles, and meetings with key people as a result of your Societies efforts. All airboat drivers deserve a special "THANK YOU" for your dedication and contribution to each field day. We could not have completed these workshops without you. Additionally, thank you to all agencies and private companies who allowed their employees to attend and bring airboats. Also, thank you to Camp Mack, they were great hosts and we appreciate the use their wonderful facilities.

One key meeting, which took place following the workshops, was with lobbyists from two herbicide companies. These interested parties sat down and discussed how we need to approach our legislators and bills in the upcoming 1996 Session. Everyone attending the meeting felt the desperate need to have a lobbyist in place to track important bills, meet key people in Tallahassee, and inform us on what to do and when to attack during the 1996 Session. A lobbyist with the needed credentials will not come cheap and everyone needs to be aware of this. However, there is a chance of splitting the cost between the Society and some herbicide companies. The need to include additional exotic plant funding for control and research was discussed to bring in larger support from other concerned areas for our bill. As current President of your Society I feel that we must support a lobbyist in Tallahassee and expand our efforts to include additional exotic plant management and research funding in 1996. If the Society agrees, now is when we need to start looking for the right person. A decision by the Board of Directors should be made regarding this matter by the 19th Annual Meeting at St.

Continued on page 22



The Great Northwest ·We all know hydrilla is not supposed to be here—but the menace does not. Hydrilla was detected in Washington this summer. Photo by David P. Tarver



CONTENTS

Hygrophila is Replacing Hydrilla in South Florida by David L. Sutton
Management of Aquatic Plant Communities in Rodman Reservoir from 1969-1994 by Judy Ludlow
Evaluation of Herbicides for Control of American Frogbit by Ken Langeland, Brian Smith, Neil Hill, Steve Grace, Mathew Cole, Johnny Drew and Ed Meadows

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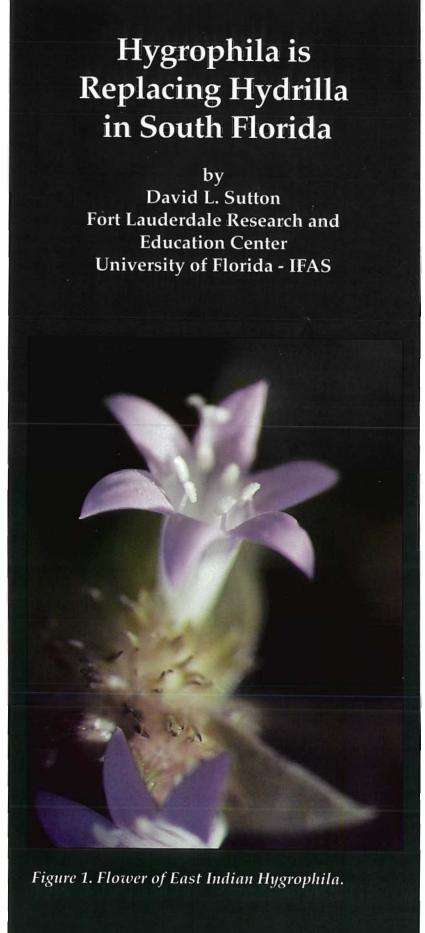


Introduction

Hydrilla (Hydrilla verticillata (L.f.) Royle) has been a serious submersed weed problem in canals and other bodies of water in the southern part of Florida ever since its introduction in the 1950's. In some locations in this part of the state Hydrilla continues to be a problem. During the past few years however, East Indian Hygrophila (Hygrophila polysperma (Roxb.) T. Anderson), is replacing Hydrilla as a major aquatic weed problem in South Florida.

Control of Hydrilla and the resistance of East Indian Hygrophila to registered aquatic herbicides are major reasons for the increase in populations of East Indian Hygrophila in South Florida. As Hydrilla has been brought under control with herbicides, open areas in the water column provide space for other plants to grow. East Indian Hygrophila now occupies many areas formerly populated with Hydrilla.

East Indian
Hygrophila was
introduced in
Florida about the
same time as
Hydrilla. It is not
known when East
Indian Hygrophila



naturalized in Florida, but it began creating problems around 1980 in several locations in Miramar and the City of Margate in South Florida. During the last few years East Indian Hygrophila has increased to the point where it is now causing serious weed problems in many locations. Little information is available on causes for the increase in growth of East Indian Hygrophila. This article will examine characteristics of East Indian Hygrophila and attempts to control its growth.

Hygrophila is in the Family Acanthaceae

The genus Hygrophila is in the family Acanthaceae. This family is a large pantropical group of herbaceous plants of 256 genera and more than 2,700 species. Plants in the family typically contain cystoliths that are evident as short, raised lines or dots in their leaves and stems. Cystoliths are accumulations of calcium carbonates formed in modified epidermal cell walls. Other family characteristics include conspicuous subtending floral bracts; two-lipped corollas (petals) associated with ovaries divided into two compart-

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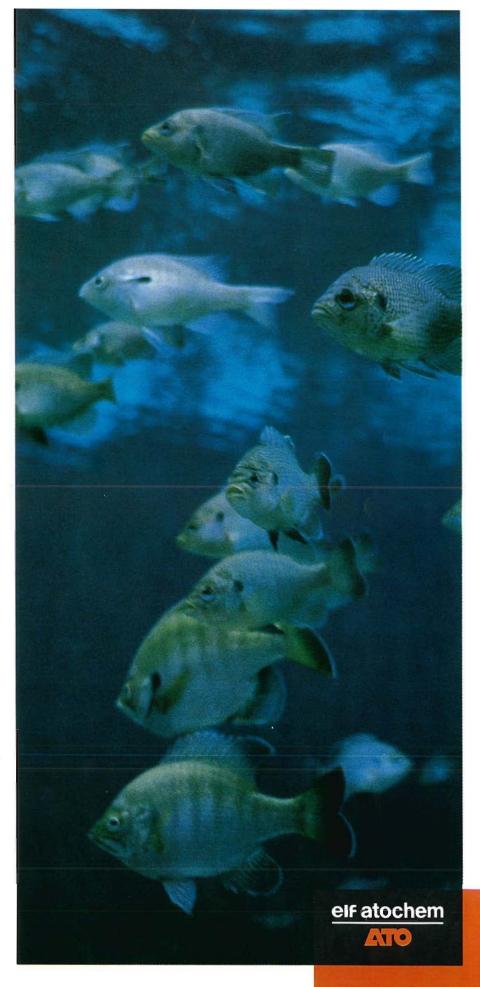
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ments; bisexual flowers; two-valved, elastically dehiscing capsules; and mucilaginous seeds with a specialized curved supporting stalk.

The aquatic genera in the Acanthaceae family are *Hygrophila* and *Justicia*. Little information is available about these aquatic genera, and no monograph has been written on *Hygrophila*. Estimates for the number of species of *Hygrophila* range from 25 to 80. More information is needed to determine the exact number of species associated with the various genera in the Acanthaceae family.

Hygrophila is derived from Greek 'hygros' and Latin '-phila' which literally means plants attracted to, or living in, or growing by preference in wet areas.

Hygrophila in Florida

Two species of *Hygrophila* occur in Florida. Lake Hygrophila (*Hygrophila lacustris* (Schlecht & Cham.)) is native to North America and occurs sporadically from northeast Florida west to east Texas. East Indian Hygrophila is native to the East Indies and was introduced into Florida around 1950 for use in the aquatic ornamental industry.

Naturalized populations of East Indian Hygrophila occur in many canals and static bodies of water in South Florida. Also, it is well established in riverine habits in Lee County on the west coast of Florida. East Indian Hygrophila appears to be sensitive to cold weather and may not survive the cool winter temperatures of central and northern Florida. However, East Indian Hygrophila has been reported to grow as far north as Virginia, but little information is available as to whether it has naturalized there or in other locations with cold winter temperatures.

Characteristics of Lake and East Indian Hygrophila

Lake Hygrophila and East Indian Hygrophila can be easily distinguished from each other by growth habit and size. Lake Hygrophila grows primarily as a marsh herb, and is an erect emergent plant

frequently reaching more than 20 inches (50 cm) in height. East Indian Hygrophila on the other hand prefers riverine habits and grows primarily as a rooted submersed plant, but it also produces emergent shoots that rarely grow more than 4 inches (10 cm) in height. Emergent shoots of East Indian Hygrophila may be present on plants growing near or on shore, and on plants in floating mats.

Both Lake Hygrophila and East Indian Hygrophila exhibit similar anatomical characteristics such as glabrous or sparsely pubescent and mostly quadrangular stems with abundant cystoliths. The stems are bluish white and dries to a yellowish color. Each flower may produce 20 to 30 seeds, but it is unknown whether the seeds are a major factor in the reproduction and spread of either species.

Growth Characteristics of East Indian Hygrophila

East Indian Hygrophila forms dense stands that occupy the entire water column (Figure 2), but it does not produce the extensive branching just below the surface of the water like Hydrilla. The stems of East Indian Hygrophila are brittle and fragment easily. These fragments form large floating mats that collect at culverts, pump stations, and



Figure 2. East Indian Hygrophila forms dense stand of submersed plants and emergent shoots above the water as shown in this canal in South Florida.

decumbent below the surface and root at the nodes. Their leaves are lanceolate or lance-elliptic and elongate-narrowed to a petiolelike base with an acute apex. The leaves of Lake Hygrophila are 2 to 4.7 inches (5 to 12 cm) in length while the leaves of East Indian Hygrophila are 0.8 to 3.1 inches (2 to 8 cm) in length.

Flowers of East Indian Hygrophila are mostly hidden in crowded apical leaf axils (Figure 1) while those of Lake Hygrophila are well distributed in distinct axillary clusters. The corolla for both species, which detaches easily, is other water control structures, and interfere with their function. Also, these fragments may be a major source of vegetative material to infest an area.

Adventitious roots form readily at the nodes of East Indian Hygrophila which aids in the rooting of fragments. Also there are reports even the leaves may root and produce new plants, but it is unknown to what extent this form of vegetative reproduction plays in the spread of this plant.

Leaves of East Indian Hygrophila above or below the surface are

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similar in shape but leaves under the water are generally larger than the emergent ones. Some aquatic plants have differently shaped leaves on the same plant depending on whether the leaves are submersed or emersed. When a plant has two or more distinct leaf shapes present on a single individual, these leaves are termed "heterophyllous." Heterophyllous leaves may differ markedly in shape, yet have similar gross anatomical organization. For East Indian Hygrophila, the significance of the similar shape of leaves above and below water on growth and development of these plants are unknown.

Flowers may be found on emersed stems of East Indian Hygrophila starting in late October and continuing on until late February. The flowers are probably self-pollinating because most set seed. The role seeds play in the spread and regrowth of East Indian Hygrophila is not known.

Water flow may play a major role in growth of East Indian

Hygrophila. Studies in small tanks in Gainesville showed that turn over rates of 1 to 5 hours produced a 3- to 5-fold increase in growth of East Indian Hygrophila. Physiological studies suggest that East Indian Hygrophila grows best at a pH of 5 to 7, and begins to photosynthesize at low light levels similar to Hydrilla.

Control of East Indian Hygrophila

East Indian Hygrophila is more resistant to currently registered aquatic herbicides than Hydrilla, and is difficult to control in canals. Summer applications of endothall at label rates to control East Indian Hygrophila in a canal in South Florida resulted in control in portions of the treatment plots for up to 12 weeks after application of herbicide, but in other areas of the canal, the plants quickly regrew to pretreatment levels within this time. The cause for this difference in plant response to application of herbicide was not readily apparent in the

study. Because endothall is rapidly degraded by microbial activity at high water temperatures of 30 C (86 F), a winter treatment with endothall was conducted to determine if control could be achieved during the cooler months of the year when a lower microbial activity would be expected.

Results from a winter application of endothall at label rates were essentially the same as for the summer application. However, dissolved oxygen was less affected by the winter treatment than the summer treatment. In both the summer and winter treatment, endothall resulted in excellent control of East Indian Hygrophila for 4 to 8 weeks followed by rapid regrowth. Additional applications approximately 8 to 10 weeks after the initial application of endothall may be required to keep East Indian Hygrophila under control.

Grass carp (Ctenopharyngodon idella Val.) have been used to control East Indian Hygrophila. Effective use of grass carp to control this plant

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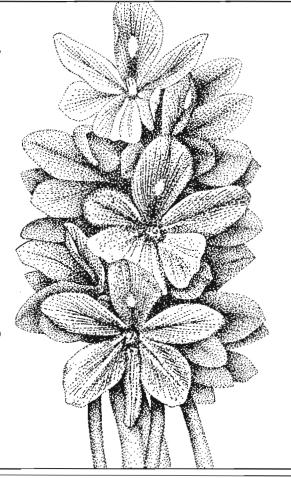
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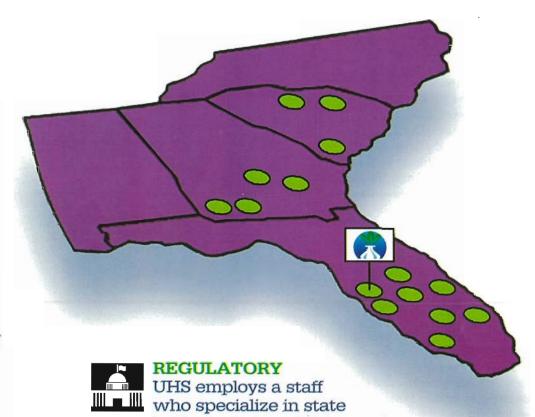
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appears to involve use of fish larger than 1.0 lb (0.45 kg) in weight, stocking rates higher than for most other weed problems, and lack of preferred plants. An aquatic weed management program using herbicides and grass carp may provide effective control of East Indian Hygrophila.

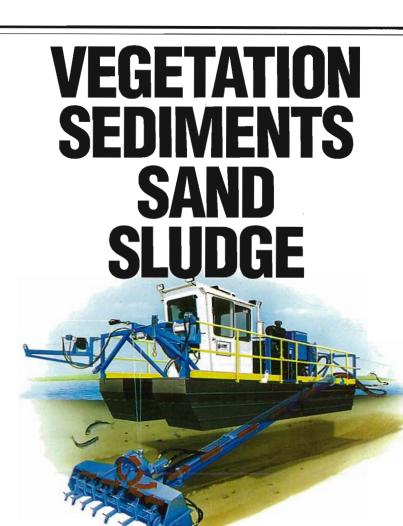
Control of East Indian
Hygrophila must also take into
account emersed plants on the
shore. Applications of Aquathol®K
liquid at a rate of 5 ounces per
gallon (39 ml per 1.0 liter) will
control shoreline emersed plants.
However, emersed plants on
floating mats are difficult to control.

Mechanical control may be useful in removing floating mats of East Indian Hygrophila. However, since the stems fragment easily, mechanical control methods need to take into account the possibility of an increase in number of these fragments that may serve as a possible source of vegetative material to infest new areas.

Solutions for long term control of East Indian Hygrophila in South Florida are not readily apparent. Effective methods are needed to prevent the spread and rapid regrowth of these plants. Aquatic plant managers need additional information to help with the control of this aquatic weed problem in South Florida.

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Management of Aquatic Plant Communities in Rodman Reservoir from 1969-1994

by Judy Ludlow Department of Environmental Protection Bureau of Aquatic Plant Management

Introduction

Rodman Reservoir (Lake Ocklawaha) was created in 1968 as part of the Cross Florida Barge Canal Project (CFBC). Since the deauthorization of the CFBC in 1991, there has been heated debate over whether Rodman should be maintained as a reservoir, or drained to facilitate restoration of the Ocklawaha River. Regardless of this debate's outcome, aquatic plant management has been, and probably will continue to be, an integral component of this system. Rodman Reservoir is presently one of only a few water bodies in Florida where effective drawdowns

can occur on a regular basis, and the ability to conduct drawdowns in Rodman Reservoir has been an important aquatic plant management tool.

The Aquatic Plant Community of Rodman Reservoir

The fertile, shallow waters of this flooded timber reservoir support a diverse assemblage of aquatic plant species. Until 1973, the dominant submerged plant species on Rodman Reservoir were southern naiad (Najas guadalupensis), Brazilian elodea (Egeria densa), and coontail (Ceratophyllum demersum), and the dominant floating plant on the reservoir was waterhyacinth (Eichhornia crassipes).

Hydrilla (Hydrilla verticillata) was first noted in the reservoir in 1971, and by 1974 had become the dominant submerged species (U.S. Army Corps of Engineers, Jacksonville District, Jacksonville 1989). By 1972, species such as waterlettuce (Pistia stratiotes), water hemlock (Cicuta mexicana), water pennywort (Hydrocotyle spp.), and others began to create a more diverse floating plant community (USACE Jacksonville District, Jacksonville 1989).

Department of Environmental Protection aquatic plant surveys have been conducted annually on Rodman Reservoir since 1982. Up to 50 species of aquatic plants have been recorded on Rodman, covering over 60 percent of the Reservoir between 1982 and 1994. Many variables such as water quality, water levels, and watershed characteristics influence aquatic plant communities. On Rodman Reservoir, some of the variables that effect submerged aquatic plants include drawdowns, control with herbicides, and the degree of water column light attenuation, which is influenced by water color (clear during drought years, tannin-stained during periods of rain).

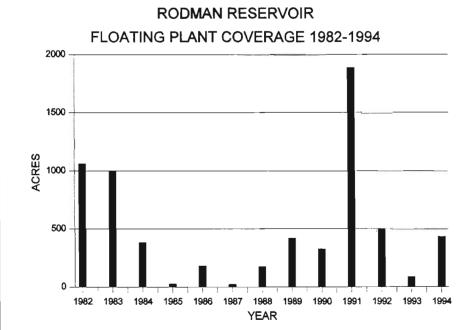
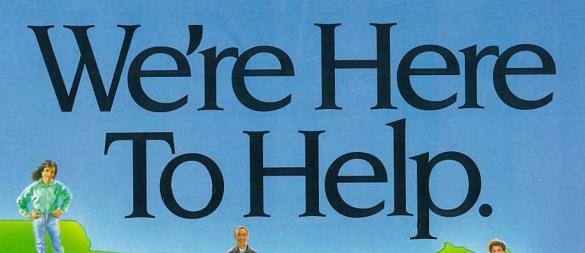


Figure 1. Department of Environmental Protection, Bureau of Aquatic Plant Management annual aquatic plant survey data for floating plants (waterhyacinth plus water lettuce) on Rodman Reservoir from 1982-1994.



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Aquatic Plant Management in Rodman Reservoir 1969-1994

Management of aquatic plants in Rodman Reservoir has generally followed the recommendations of the Cross Florida National Conservation Area Operational Management Plan (USACE Jacksonville District, Jacksonville 1989). These recommendations are 1: "Floating aquatic plants will be kept to the lowest level compatible with desirable management goals and past experience with other controlling factors." and 2. "In general, the aquatic plant community should be managed for maximum diversity and dominance by native species. The Lake Ocklawaha management plan encourages native flora and fauna by managing nearnatural hydroperiod regimen for water levels."

Many aquatic plant management programs typically use a combination of herbicides, drawdowns, and mechanical and biological controls. Although that is true for aquatic plant management in Rodman Reservoir, herbicides and drawdowns have been the most commonly used methods. On Rodman Reservoir, drawdowns have usually occurred in late summer to early fall, dropping water levels in the reservoir by 3 to 8 feet, and lasting throughout the winter. Stranded aquatic plants are killed by desiccation and also by winter frosts.

Waterhyacinth and Waterlettuce

Waterhyacinth and waterlettuce, are managed in Rodman Reservoir using the herbicides, 2,4-D and/or diquat, and reservoir drawdowns (Table 1). Floating plant populations have covered as much as 1800 acres of the reservoir, but have generally remained below 500 acres since 1982 (Figure 1).

Hydrilla

In Rodman Reservoir hydrilla is difficult to manage, but has generally remained below 3500 acres since 1982 (Figure 2). The number of submerged obstructions in this

RODMAN RESERVOIR HYDRILLA COVERAGE 1982-1994

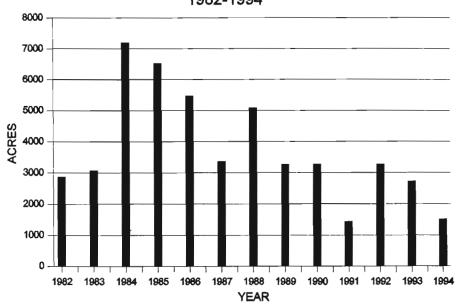


Figure 2. Department of Environmental Protection, Bureau of Aquatic Plant Management annual aquatic plant survey data for hydrilla in Rodman Reservoir, 1982-1994. Data from 1982-1992 are adjusted for a 18 ft. NGVD reservoir size of 9,600 acres.



submerged obstructions in this flooded timber reservoir make mechanical harvesting extremely difficult. Hydrilla was managed using the herbicide fluridone in 1988 and 1990 (Table 1), but because flow rates average over 1600 cubic feet per second (Rao et al. 1994), the cost effective use of herbicides is limited to protected shorelines and cove areas. Drawdowns have been used in Rodman Reservoir, with some degree of success, to control hydrilla (Haller and Shireman 1984). Also, observations from reservoir biologists indicate that during rainy years the inflow of dark, tannin-stained water suppresses the growth of hydrilla in Rodman Reservoir (Dean Barber DEP, Jim Kelley DEP, Dave Bowman USACE pers comm.)

Funding Aquatic Plant Management in Rodman Reservoir 1969-1994

Until the deauthorization of the Cross Florida Barge Canal in 1991, Rodman Reservoir was managed directly by the U.S. Army Corps of Engineers (Corps). From October 1992 to the present, aquatic plants on Rodman Reservoir have been managed through the Department of Environmental Protection, by the St. Johns River Water Management District, under the Cooperative Aquatic Plant Control Program. The Cooperative Aquatic Plant Control Program is funded by the department and the Corps for plant control in public waters.

Based on data from the Corps, Jacksonville District, and the Department of Environmental Protection, the 25 year average cost of controlling waterhyacinth, waterlettuce, and hydrilla on Rodman Reservoir from 1969 -1994 is approximately \$31,000 (Table 1). The highest costs to date were in 1970 (\$235,700) and 1971 (\$217,600) when aircraft were used to control over 4,000 acres of waterhyacinth onthe reservoir. After waterhyacinth was brought under control in the early 1970's, drawdowns and herbicides have been

used to manage floating plants and hydrilla for a 22 year average annual cost of \$14,000.

Summary

The management of aquatic plants in Rodman Reservoir is somewhat unique in Florida in that drawdowns have often been used as a management tool. The drawdowns which have occurred on the reservoir have demonstrated that. in combination with other aquatic plant management techniques, temporary control of hydrilla, waterhyacinth, and waterlettuce can be achieved while, at the same time, enhancing fish and wildlife habitat (Haller and Shireman 1984). Regardless of the outcome of the Rodman Reservoir debate, aquatic plant control will likely continue to be an important component of the management of this system.

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P.O. Box 6006 • Vero Beach, FL 32961

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U.S. Army Corps of Engineers. 1989. Cross Florida National Conservation Area Operational Management Plan (Authorized Portion), Jacksonville District, Jacksonville, Florida.

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Table 1. Summary of floating plant (waterhyacinth and waterlettuce) and hydrilla management on Rodman Reservoir from 1969 to October 1994. Data are from U.S. Army Corps of Engineers, Jacksonville District, Jacksonville (1969-1993) and Department of Environmental Protection (1993-1994). "na" = not available.

FISCAL YEAR	ACRES TREATED FLOATING HYDRILLA		METHOD OF CONTROL	APPROXIMATE COST <i>FLOATING HYDRILLA</i>			
1969-1970	144		Herbicides	\$	7,200		_
1970-1971	4714		Herbicides by Aircraft	\$:	235,700		
1971-1972	4352		Herbicides by Aircraft		217,600		
1972-1973	64		Herbicides & Drawdown	\$	3,200		
1973-1974	138		Herbicides	\$	6,900		
1974-1975	358		Drawdown	\$	17,900		
1975-1976	594		Herbicides	\$	29,700		
1976-1977	356		Herbicides	\$	17,800		
1977-1978	17		Herbicides	\$	850		
1978-1979	26	6	Herbicides	\$	1,300	na	ı
1979-1980	0		Drawdown	\$	0		
1980-1981	12		Drawdown	\$	600		
1981-1982	0	2	Drawdown	\$	0	\$	0
1982-1983	0	6	Experimental	\$	0	na	
1983-1984	0			\$	0		
1984-1985	0			\$	0		
1985-1986	0		Drawdown	\$	0		
1986-1987	0	6	Herbicides	\$	0	\$	1,200
1987-1988	71		Herbicides	\$	10,650		
1988-1989	40	222	Herbicides & Drawdown	\$	6,000	\$	87,600
1989-1990	184		Herbicides	\$	27,450		
1990-1991	225	63	Herbicides	\$	33,750	\$	36,400
1991-1992	176		Herbicides & Drawdown	\$	26,400		
1992-1993	33		Herbicides	\$	4,950		
1993-1994	14	3	Herbicides	\$	2,100	\$	1,800
TOTAL FLO	ATING PLANT	MANAGEMENT	COSTS 1969-1994	\$	650,050		
TOTAL HYD	ORILLA MANAC	EMENT COSTS	1969-1994			\$	127,000

1995 FAPMS Annual Meeting



The Florida Aquatic Plant Management Society will hold it's 1995 Annual Meeting at the St. Petersburg Hilton, which is located at 333 1st Street South, St. Petersburg. The Society has blocked 200 rooms on the evenings of October 17, 18, and 19. The contract rate for the rooms is \$59.00 single or double pe night. The room block will be held until 30 days prior to the meeting, or until September 16, 1995. The hotel offers us a large parking lot next to the building for our equipment demonstration. St. Petersburg is a lovely town and the hotel is located on the bay very near to the municipal pier, so there will be shopping and restaurants close by.

Meet in St. Pete!

For registration information contact Nancy Allen, Secretary 904/795-2239, FAX 904/795-1911.



Evaluation of Herbicides for Control of American Frogbit

by Ken Langeland, Brian Smith, Neil Hill, Steve Grace

University of Florida IFAS, Agronomy, Center for Aquatic Plants Gainesville, Florida

Mathew Cole, Johnny Drew, Ed Meadows St. Johns River Water Management District Palatka, Florida

Introduction

American frogbit (Limnobium spongia) is a native rooted or floating aquatic plant found in still or slow moving waterways such as ponds, lakes and canals. Because American frogbit is a native plant, it is often considered a non-target species or of minor importance in aquatic plant management programs. However, frogbit populations can become expansive, and cause problems similar to other floating plants, such as waterhyacinth and waterlettuce. For example, during June through July 1992, large floating mats of American frogbit, mixed with hydrilla, broke loose from Lakes Sawgrass, Little Sawgrass, and Hell'n Blazes, in the upper St. Johns River, and caused severe blockages at the SR 192 bridge (Figure 1). Removal of these blockages by mechanical means was necessary at a cost of over \$60,000. Similarly, floating mats of American frogbit, when allowed to reach large population levels in Upper Myakka Lake, have broken loose during high flows and required costly mechanical removal at bridges, by the Southwest Florida Water Management District (Figure 2).

American frogbit has been reported as difficult to control with herbicides, and aquatic plant

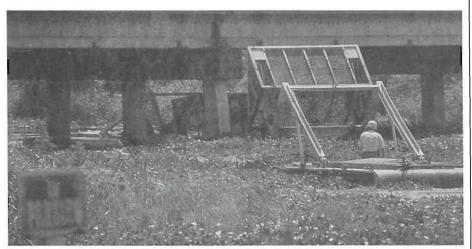


Figure 1. South Florida Water Management District towboats assist the St., Johns River Water Management District in removing a jam of American frogbit and hydrilla from the SR 192 bridge over the St. Johns River (photo by Wayne Corbin).

managers in Florida differ in their opinions on the best control method. Eddie Knight said that "Frog's-bit is probably the most difficult emergent plant to control," but could be controlled with a tankmix of four quarts of 2,4-D, two quarts of diquat, and 3/4 pint X-77® per acre, as long as coverage is thorough (Aquatics, March 1985). Others say (personal communications) they use two quarts of diquat with one pint of 2,4-D, while some say they use two quarts or less of diquat alone (along with an appropriate surfactant). It has been reported that regrowth occurs after any of these applications.

The following studies were

conducted to 1) evaluate efficacy of diquat, 2,4-D amine, and tankmixes of the two herbicides 2) determine the importance of diluent volume with diquat and 2,4-D, and 3) evaluate glyphosate in combination with two different surfactants (Kinetic® and Induce®) for American frogbit control.

Materials and Methods
Evaluation of 2,4-D amine and
diquat on the control of American
frogbit. American frogbit plants,
collected from a local lake, were
placed uniformly in 100 ft² (9.5 m²)
floating PVC quadrats in three 0.24
acre (0.1 ha) ponds (replicates).
Plants were allowed to acclimate for





four weeks, after which foliar applications of 2,4-D amine (3.8 lb ae/gal) and diquat (2.0 lb cation/gal) were applied at 0.25, 1.0, 2.0, and 4.0 qt/ac and 0.25, 1.0, and 2.0 qt/ac, respectively, and in combination. An organosilicone surfactant (Kinetic®) was added to the diluent volume at 0.25% v/v. Treatments were applied May 18, 1993 using a portable electric sprayer calibrated to deliver 200 GPA (1890 l ha⁻¹). Well water was used as the diluent.

Visual evaluations (not reported) were made once a week for four weeks and biomass was evaluated from a 1/4-m quadrat four weeks post treatment. Control was evaluated as reduction in biomass compared to untreated checks and expressed as a percent.

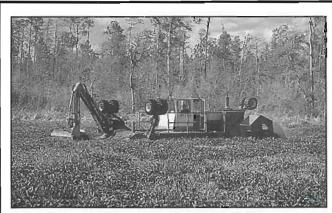
Influence of diluent volume on diquat and 2,4-D amine efficacy on American frogbit. Research was conducted in conjunction with the St. Johns River Water Management District on a natural population of American frogbit growing in



Figure 2. Floating mats of American frogbit and associated vegetation, such as this one, when allowed to reach large population levels in Upper Myakka Lake, have broken loose during high flows and required costly mechanical removal at bridges, by the Southwest Florida Water Management District.

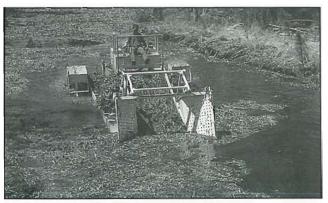
the littoral zone of Lake Lochloosa, Alachua County, Florida. Floating PVC quadrats, of 25 ft² (2.4 m²), were placed over actively growing plants in an arrangement for three replicates of each herbicide/diluent treatment. Foliar applications of diquat (2.0 lb cation/gal) were applied at 1.0 and 2.0 qt/ac and

tank-mixed with 0 or 2.0 qt/ac 2,4-D amine. An organosilicone surfactant (Kinetic®) was added to the diluent volume at 0.25% v/v. Diluent volume was calibrated at 100 or 200 GPA (945 or 1890 l/ha, respectively) using a portable electric sprayer. Lake water was used as the diluent.



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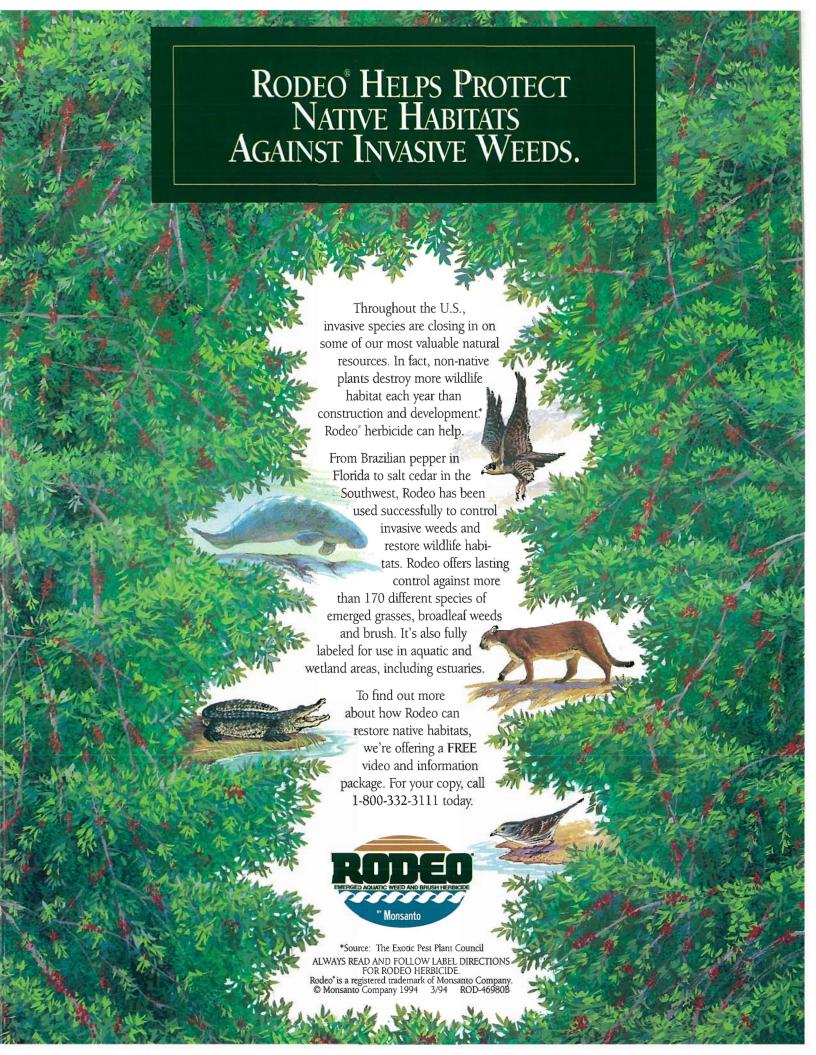
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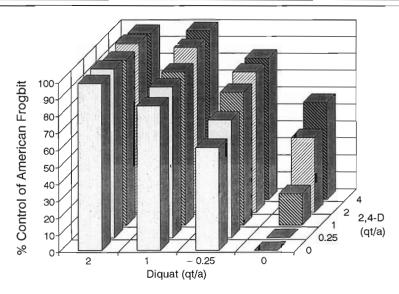


Figure 3. Control (% redection in biomass compared to untreated checks) of American frogbit observed four weeks after application of various rates of diquat, 2,4-D, or combinations of the two herbicides.

Visual evaluations, which compared discoloration and necrosis of treated plants to untreated checks, were made weekly for four weeks. A scale of 0% to 100%, with 0% representing no discoloration or necrosis and 100% meaning that all plants were dead was used.

Evaluation of glyphosate for American frogbit control. This experiment was conducted on a natural population of American frogbit, which had entirely covered the surface of a small lake in Suwanee County, Florida. Floating PVC guadrats, of 100 ft2 (9.5 m2), were placed over actively growing plants in an arrangement for three replicates of each herbicide/surfactant treatment. After placement, the PVC quadrats were anchored in place. Glyphosate (4.0 lb ae/gal) was applied to the foliage at 0, 1.0, 2.0, and 4.0 qts/ac in combination with either an organosilicone surfactant (Kinetic®) at 0.25% v/v or a nonionic surfactant (Induce®) at 0.5% v/v. All treatments were applied July 6, 1992 using a portable electric sprayer calibrated to deliver 150 GPA (1420 l ha-1). Lake water was used as the diluent.

Visual evaluations were used to compare treatments as described for the spray volume study.

Results and Discussion Evaluation of 2,4-D amine and diquat on the control of American frogbit. Diquat at 1 or 2 qt/ac, either alone or in combination with any rate of 2,4-D, provided excellent control (88-95%) of American frogbit (Figure 3). Therefore, addition of 2,4-D when using these rates of diquat in 200 GPA spray volume does not appear to provide a control advantage. 2,4-D alone provided less than 60% control at any of the rates tested. Tank-mixing 2,4-D with the lowest diquat rate (0.25 qts/ac) increased activity, but did not provide as good control as the higher rates of diquat alone.

The influence of diluent volume on diquat and 2,4-D efficacy on American frogbit. When diquat was applied at 1 qt/ac, doubling the

diluent volume from 100 to 200 GPA markedly improved frogbit control from <40% to >90% (Figure 4). Likewise, when 2 qts/ac 2,4-D was tank-mixed with 1 qt/ac diquat, doubling the diluent volume from 100 to 200 GPA improved frogbit control from <50% to >90%. Increasing the diluent volume when diquat was applied at 2 qts/ac alone or tank-mixed with 2,4-D did not statistically improve frogbit control. Tank-mixing 2,4-D with diquat did not significantly improve frogbit control at comparable diquat rates.

Evaluation of glyphosate for American frogbit control. Plants treated with glyphosate and Kinetic® demonstrated greater discoloration than plants treated with Induce® and glyphosate at equivalent glyphosate rates (Figure 5). However, glyphosate did not provide adequate control of American frogbit at any of the rates tested or with either of the surfactants tested (Figure 5). Discoloration or patchy necrotic lesions were observed, but plants were observed to be fully recovered on site visits two and three months post treatment.

Summary

Results of these studies agree with aquatic plant managers who have reported control of American frogbit with diquat or diquat and 2,4-D combinations. The control observed in this study with the combination of 0.25 qt/ac diquat

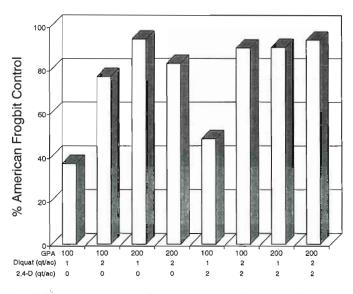


Figure 4. Control (visual evaluations) of American frogbit four weeks after application of diquat, 2,4-D, and both herbicides together, at two different diluent volumes.

and 2.0 qt/ac 2,4-D suggest that less diquat may be necessary when combined with 2,4-D, but this interpretation requires further field testing. A benefit was not observed of adding 2,4-D when 1.0 or more qt/ac diquat was used. However, under conditions that were not tested in this study, addition of 2,4-D may enhance control.

Adequate coverage of foliage is necessary when using a contact type of herbicide such as diquat. Inadequate control of American frogbit with diquat or diquat and 2,4-D reported by some aquatic plant managers may be related to poor coverage. When recommending 2,4-D and diquat for American frogbit control, Eddie Knight cautioned that "-good coverage of the solution is important. The extra time spent soaking the plants normally insures a good kill-." Our data supports Mr. Knight's contention because we found 200 gallons per acre of spray-volume necessary for adequate coverage. We recommend using spray vol-

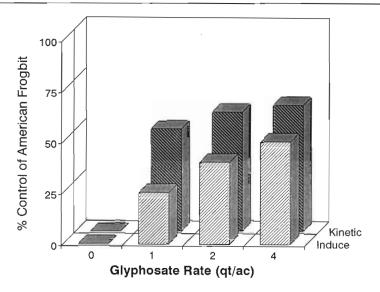


Figure 5. Control (visual observations) of American frogbit observed four weeks after application of various rates of glyphosate with two different surfactants.

umes in this range to expect comparable results in the field.

Acknowledgements

This material is based upon research supported in part by IFAS/ARS cooperative agreement No. 58-XY-9-001 and IFAS/SJRWMD cooperative agreement No. 94D270.

We appreciate the support of the following companies for providing herbicide formulations or adjuvants for the studies: DowElanco, Helena Chemical Company, Monsanto Company, Rhone-Poulenc Ag Company, Valent U.S.A, Zeneca Professional Products Corporation. Published as IFAS Circular 1158.

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AQUAVINE



Yeah We're Moving Up

Mathew Cole, former Technician Supervisor for the SJRWMD in Palatka, has accepted a position with Environmental Waterways, Inc. Matt will be reunited with his former St. Johns boat partner, Darryl Blackall, who is the office manager of the Orange Park Office. Johnnie Drew, 6-year veteran of the hydrilla wars with St. Johns, has been appointed interim supervisor. Johnie's first comment was, "Hey Wayne how do you calibrate this 486 DX —?"

Welcome Randall Stocker Randall Stocker is now the Director for the UF/IFAS Center for Aquatic Plants in Gainesville. Randall will be wanting to get out and about to meet everyone, so give him a call at 904/392-6841 and invite him to your place.

Meetings

South Carolina Aquatic Plant Management Society Annual Meeting, Springmaid Beach Recreation and Conference Center in Myrtle Beach, October 4-6, 1995 (postponed due to Felix). Contact Larry McCord 803/761-8000 ext. 5735.

Florida Aquatic Plant Management Society Annual Meeting, St. Petersburg Hilton, October 17-19, 1995. Contact Mike Hulon, President 407/ 846-5304.

Southern Weed Science Society Annual Meeting, Adams Mark Hotel, Charlotte, NC. January 15-17, 1996. Contact Barry Sims 901 / 855-1472.

Weed Science Society of America 1996 Annual Meeting, Norfolk, Virginia Marriot and Omni Hotels, February 6-9, 1996. Contact Dr. Henry P. Wilson 804/442-6411.

Weed Science Society of America 1997 Annual Meeting, Orlando, FL, Clarion Hotel, February 2-6, 1997.

Letter

Continued from page 3

Petersburg in October. We will announce our decision at the annual meeting.

The Department of Environmental Protection funding for hydrilla control in fiscal Year 1995-96 was increased by our State Legislators and totals 5.8 million dollars. This money is greatly appreciated; however, it runs out in FY 1996-97 and all that remains for statewide hydrilla control is \$300,000. Everything we have done to date will be lost if there is not a dedicated funding source found soon for hydrilla management.

For more information or to express your opinion on these issues, please contact me at 407-846-5300.

Thank you, Mike Hulon, President

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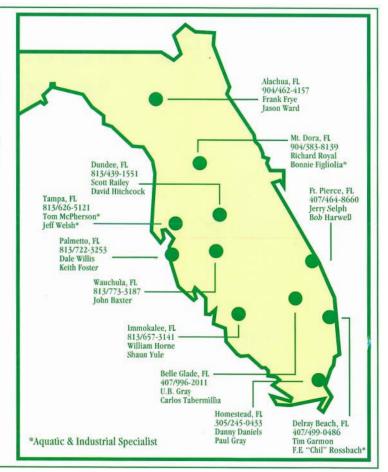
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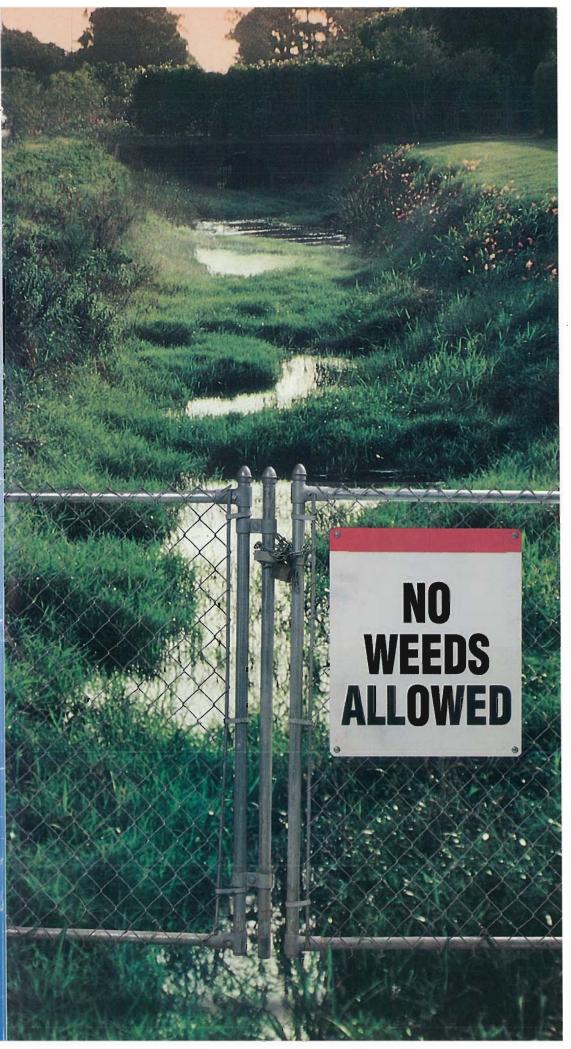
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